

# ivgN EXPLORING THE ROLE OF STEM CELLS IN CANCER THERAPY

Swaha Bhattacharya<sup>1</sup>, Sandeep Poddar<sup>2\*</sup>

<sup>1</sup> Post Graduate Department of Biotechnology, St. Xavier's College, Mother Teresa Sarani, Kolkata 700016, India  
Email: simplyswaha@rediffmail.com

<sup>2</sup> Lincoln College, Mayang Plaza, Block A, No. 1 Jalan SS26/2, Taman Mayang Jaya, 47301 Petaling Jaya, Selangor Darul Ehsan, Malaysia. E mail: sandeep@lincoln.edu.my

\*Corresponding author

## ABSTRACT

The conventional methods for cancer treatment used so far are radiotherapy and chemotherapy. But, the drawbacks of these procedures are that most chemotherapy drugs and radiation techniques target the normally growing healthy cells of the body. They target the normal tissue as well, while being unable to completely destroy the cancer stem cells which are highly resistant to these techniques. Solid tumors such as breast cancer and colon cancer possess a minor population of tumorigenic cells termed cancer stem cells (CSCs) or tumor initiating cells (TICs). A recent approach to deal with these problems is by making use of stem cells. Stem cells are generally defined by an ability to differentiate into multiple cell lineages and self-renew. These stem cells have a number of utilities. They contribute to not only organogenesis but also help in regeneration in response to the injury of tissues and organs. Several studies conducted in the biology of stem cells have allowed for their identification and characterization in a variety of tissues and organs. This review makes an attempt to explore the behaviour and potential of stem cells in cancer treatment alongside the traditional methods of chemotherapy, radiation therapy and surgical removal of tumour.

*Key words:* Cancer, Stem cell, Therapy

## INTRODUCTION

For a considerable period of time now, cancer has been causing distress and death in the world. Environmental stress and increasing amount of carcinogens circulating freely in the environment due to technological advances strangled hold of cancer. Despite recent advances in the treatments of cancer, the clinical outcome is yet far from expectation. The conventional methods for cancer treatment used so far are radiotherapy and chemotherapy. But, the drawbacks of these procedures are that they affect both normal and cancerous tissue while being unable to completely destroy the cancer stem cells which are highly resistant to these techniques (Moserle et al. 2010). A recent approach to deal with these problems is by making use of the array of cells, the stem cells. Stem cells are generally defined by an ability to differentiate into multiple cell lineages and self-

renew. They contribute to not only organogenesis but also regeneration in response to the injury of tissues and organs (Millera et al. 2005). Advanced studies in the biology of stem cells have allowed for their identification and characterization in a variety of tissues and organs. It has also been documented that in solid tumors such as breast cancer and colon cancer, there exists a minor population of tumorigenic cells which can generate new tumors in xenograft transplantation (Houghton et al. 2009). Cancer stem cells (CSCs) or tumor initiating cells (TICs) can be defined as cells with a tumour initiating potential (Wang & John 2005). They comprise a very small population in tumours and are known as tumorigenic cells, or tumor-initiating cells (TICs) (Chiba et al. 2009). These cancer stem cells possess stem cell-like properties, contributing to a hierarchical structure containing varied progenies in a similar fashion to normal stem cells.

Normal stem cells are characterized by three main properties:-

- (1) the presence of an extensive capacity for self-renewal, which allows maintenance of the undifferentiated stem cell pool over the lifetime of the host;
- (2) strict regulation of stem cell number; and
- (3) the ability to undergo a broad range of differentiation events to clonally reconstitute all of the functional elements in the tissue (*Clarke 2005*).

Two conceptual models of tumor growth have been proposed (*Martin et al. 2008, Krishnamurthy et al. 2004*). In the older, stochastic model, all cells in the tumor have a high proliferative capacity. A growth fraction of <1 still occurs due to individual cell loss and non-reproduction, the result of constraints of the microenvironment such as focal lack of nutrients or oxygen, and age. However, in many cancers a stem cell model of tumor reproduction probably takes place. As in normal tissues, only a small percentage of the tumor population (i.e., the CSCs) maintain the capacity for long-term proliferation, while most cells proceed forward in a process of aberrant terminal differentiation.

Stem cells can be divided into three main categories: embryonic, germinal, and somatic. Embryonic stem cells (ESCs) originate from the inner cell mass of the blastocyst. ESCs are pluripotent and have indefinite replicative life span, which can be attributed to their telomerase expression. Germinal stem cells are derived from primary germinal layers of embryo. They differentiate into progenitor cells to produce specific organ cells. Somatic/adult stem cells (ASCs) are progenitor cells as they are less totipotent i.e. have less replicative life span than ESCs. They exist in mature tissues such as haematopoietic, neural, gastrointestinal and mesenchymal tissues. ESCs are derived from 5-day old pre-implantation human embryos while ASCs can be obtained from a number of tissues like bone, synovium, brain, adipose tissue, blood vessels, blood and umbilical cord blood. Owing to reasons that are both ethical and legal, the use of ESCs is restricted in research and clinical fields, making ASCs the main supplement for the stem cells. More than two thirds of cancer patients currently receive either chemotherapy or radiotherapy, which in many

cases have beneficial effects and can improve quality of life and survival. In spite of the broad use of these therapies, there still remain many unresolved issues regarding the mechanisms that determine remission or the subsequent relapse of the tumor.

## OBJECTIVE

The objective of this present review is to understand the role of stem cells in the treatment of cancer. This study explores the problems associated with the conventional therapies of cancer and the prospects of stem cells as a complementary therapy in addition to the traditional techniques of chemotherapy, radiotherapy and surgery. The origin, characteristic and application of stem cells has been highlighted.

## STEM CELLS AND CANCER

The subject of stem cells in malignancy is one that still remains obscure today as compared with their role in normal tissue turnover. Tumors are composed of a heterogeneous mixture of tumor cells at various levels of differentiation. Only a small population of cells within a tumor possess self-renewal capacity (*Daniel & Brunschwig 1961, Harrington 2004*). Several studies have suggested, that while all cells within a tumor are equal, at any given time, only a small fraction of cells is in an appropriate state or is stimulated by appropriate external signals to form a new tumor. On one side, a predetermined population of cells exists with the "cancer stem cell" phenotype, enabling this cell to perpetuate the tumor, while on the other side, cells of the same tumor are incapable of self renewal. In order to prove the latter, prospective isolation of this population is required. John Dick and his group demonstrated that cells capable of establishing a human AML phenotype in a recipient mouse were isolated only within the cell fraction expected to contain the hematopoietic stem cells which is defined by the CD34<sup>+</sup> CD38<sup>-</sup> phenotype (*Vormoor et al. 1994*). These cells could be passed from animal to animal and maintain the AML phenotype (*Dick 1996*), confirming the property of self-renewal. As a result for the first time, it was triumphantly demonstrated that there are cells within the tumor which have properties similar to stem cells, i.e., they possess the capacity to reconstitute the tumor when transplanted into an

appropriate recipient (differentiation) through several rounds of transplantaion (self-renewal). A similar approach has led to the identification of subpopulations of tumor cells with stem cell properties within breast tumors (*Al-Hajj et al. 2003*), gliomas (*Singh et al. 2003*, *Singh et al. 2004*, *Sanai et al. 2005*), melanoma (*Fang et al. 2005*), prostate cancer (*Collins et al. 2005*) and osteosarcoma (*Gibbs et al. 2005*). Thus, the result of these observations is the "cancer stem cell hypothesis" (*Reya et al. 2001*). This hypothesis postulates that within a tumor, a small proportion of cells with unlimited proliferative capacity drive tumor growth. The observation that most cancers comprise a heterogeneous population of cells that have undergone varying degrees of differentiation leads to this model. The origin of CSCs has been quite obscure. For a long time, there has been conflicting beliefs of CSCs originating from normal (somatic) stem cells or from non-stem progenitors or differentiating cells. However, recently conducted studies that have been emphasize a progenitor cell origin for many types of leukaemic stem cells along with the stem cell origin (*Lobo et al. 2007*). In solid tumors too, it is most likely that not only somatic stem cells but also differentiating progenitor cells are capable of becoming CSCs (*Yang & Chang 2008*). Rapp et al. proposed a model of oncogene-induced plasticity for CSC origin by demonstrating reprogramming events triggered by a specific combination of oncogenes. *Li et al. 2009*, suggested that genomic instability is a driving force for transforming normal stem cells to CSCs and, in CSCs, a potential mechanism for cancer cell heterogeneity.

### **CANCER STEM CELL NICHE**

The concept of stem cells existing in a unique microenvironment was first proposed by *Schofield 1978* (*Schofield 1978*). The stem cell niche comprises the microenvironment surrounding normal and cancer stem cells. It plays several roles including providing a mechanical anchorage for the stem cells and in cross-talk communication mediated by direct contact and/or indirect extracellular factors. Wnt ligands are produced and released from both stem cells and niche cells, BMP and Sonic hedgehog (Shh) are released from niche cells and epithelial cells respectively, and Notch signaling is transmitted between neighboring cells. The microenvironment may also provide signaling via the

cell receptor integrin as suggested by its expression in prostatic CSCs (*Collins et al. 2005*) and its co-expression with AC133 (CD133) in the epidermal basal cells (*Yu et al. 2002*). The concept of a CSC niche is a matter of debate (*Li et al. 2009*). Several possible models were proposed to clear this debate (*Sneddon & Werb 2007*). CSCs are capable of surviving the normal stem cell niche, a distinct CSC niche is necessary for activation, CSCs may be capable of providing signals that instruct an otherwise quiescent niche to become activated ("hijacking the niche"), CSCs could amplify an already existent activated niche, CSCs may be niche-independent, that is, they themselves acquire the ability to maintain its activity, and there may be a discrete niche that is inhibitory for CSC maintenance. No particular single model fits all the different types of cancer. Further research is required to set up a CSC niche theory that can be universally accepted.

### **CANCER STEM CELL MARKERS**

Several specific markers are utilized to detect and isolate CSCs from among the innumerable cancer cells and stromal cells occupying the entire tumor tissue (*Yang & Chang 2008*, *Hirschmann-Jax et al. 2004*). This is primarily because CSCs constitute a very small fraction of the cancer cell population. Some of the markers include CD34 in several kinds of leukemia and CD44 in breast cancers, colorectal cancers, pancreas cancers, prostate cancers, head/neck cancers and some bone sarcomas.

Some of the important cancer stem cell markers are elaborated below.

#### **CD34**

CD34 was reported to be expressed in common acute lymphoblastic leukemia cells in 1987. It was suggested that CD34-positive leukemic cells may represent a less differentiated phenotype form than CD34-negative ones (*Ryan et al. 1987*). CD34 which is a heavily glycosylated type I transmembrane molecule can be phosphorylated by a variety of protein and tyrosine kinases. It is confirmed to be a sialomucin (*Lanza et al. 2001*). In contrast to the high endothelial venules for which CD34 serves as a ligand for 1-selectin, CD34 is not the ligand for 1-selectin in hematopoietic stem/progenitor cells. Ligands for hematopoietic CD34 remain to be identified (*Lanza et al. 2001*).

### CD44

CD44 which is originally described as a leukocyte-homing receptor, is made up of a family of glycoproteins encoded by a single gene, which vary in size due to alternative splicing. It functions as a pleiotropic factor and is important for tissue remodeling, cell-to-extracellular matrix adhesion, and cell migration. CD44 is the major receptor of hyaluronan (previously known as glycosaminoglycans), carbohydrate polymers of the extracellular matrix (Stern 2008). CD44 has been utilized as a CSC marker not only for leukemia but also for a variety of solid cancers.

### CD133

A broad spectrum of malignant tumors including brain tumors, colorectal cancers, pancreatic cancers, breast cancers, prostate cancers, ovarian cancers and some lung cancers have CD133 as a specific marker of CSCs (Eramo et al. 2008). CD133 was first reported as a novel marker for human hematopoietic stem and progenitor cells (Yin et al. 1997). It was later found in some types of leukemic cells (Martin et al. 2008). CD133 expression has been detected in human central nervous system stem cells (Uchida et al. 2000), human trophoblasts (Potgens et al. 2001), human lymphatic/vascular endothelial precursor cells (Salven et al. 2003), and human prostatic epithelial stem cells (Richardson et al. 2004). The CD133 antigen is a 120 kDa five-transmembrane domain glycoprotein, and its chromosomal location (4p16.2-p12) and amino acid sequence have been resolved (Piechaczek 2001). CD133 is actually detected by its glycosylated epitope, AC133, and it is likely that AC133, not CD133, is a more reliable CSC marker (Mizrak et al. 2008). A number of other studies have emphasized that the use of CD133 expression as a marker for CSC should be critically evaluated (Bidlingmaier et al. 2008). Such reports may explain the inconsistency observed in the results from different studies.

## RESISTANCE TO CHEMOTHERAPY

It has been suggested experimentally that side population (SP) and CSC have higher resistance to chemotherapy than the bulk of cancerous cells. Hirschmann-Jax et al. first demonstrated that neuroblastoma SP cells were less sensitive to

mitoxantrone (Hirschmann-Jax et al. 2004). Szotek et al. 2006 states doxorubicin could not inhibit the SP fraction of ovarian cancer, unlike non-SP cells showed similar results. The fact that this drug preferentially kills non-SP cells (Chua et al. 2008) is further reinforced by the fact that the SP of human malignant glioma cell lines and primary glioblastoma was shown to increase following treatment with temozolomide. A number of studies conducted on CD133+ brain tumor stem cells disclosed a survival advantage compared to CD133- cells to temozolomide, carboplatin, paclitaxel and etoposide (Eramo et al. 2008).

One key property is resistance to apoptosis (Moserle et al. 2010). Stem cells are programmed to be long-lived, in order to maintain the progenitor pool from which differentiated cells derive. For this purpose, stem cells activate some protective mechanisms that shield them from senescence and cellular stress. These mechanisms include: (I) activation of some self-renewal pathways, such as TGF- $\beta$ , Sonic Hedgehog (SHH), Wnt/ $\beta$ -cat or BMI-1; (II) expression of anti-apoptotic proteins like BCL-2; (III) enhanced capability to repair DNA damage after genotoxic stress; (IV) generation of autocrine loops through the production of growth factors like epidermal growth factor (EGF); and (V) over-expression of drug-effluxing pumps and metabolic mediators, that allow the cell to rapidly eliminate or degrade toxic compounds and radical oxygen species (Moserle et al. 2010). A point of great importance is that it appears that resistance to apoptosis, which can be limited to CSC initially, is often rapidly acquired also by the bulk of tumor cells at relapse. This might be attributed to the genetic instability which distinguishes tumor from normal cells. The result is that chemotherapy invariably causes bone marrow toxicity due to its effects on trans-amplifying, progenitor and even more differentiated cells. Although tumors may initially show regression, they inadvertently become completely resistant to chemotherapy. After apoptosis, a second key property is expression of certain pumps, namely ABCB1, ABCG2 and MDRI. These are the principal mediators of multidrug-resistance identified so far. They transport both hydrophobic and hydrophilic compounds, helping the cell to extrude several drugs including vinblastine, paclitaxel, mitoxantrone, doxorubicin, topotecan, methotrexate and imatinib mesylate (Yin et al. 1997). Although the exact physiological role of these pumps is still

under scrutiny, it is known that they are involved in cellular protection against exogenous products and in resistance to hypoxic stress, mediated by an increased ability to consume hydrogen peroxide (*Martin et al. 2008*) and a reduced accumulation of toxic heme metabolites (*Krishnamurthy et al. 2004*). In normal stem cells these pumps play a role in the repression of cellular differentiation and are generally turned off in committed progenitors and mature cells. The enhanced expression of these pumps in SP may explain their particular drug resistance and their tolerance to hypoxic conditions and as a result, make these proteins ideal targets for cancer therapy. Cancer stem-like cells were separated by the SP fraction method from hepatoblastoma cells. The in vivo experiment by *Hayashi et al. (2011)* proved that SP fraction cells inoculated into mice were self-replicated, and also identified the existence of cancer stem-like cells (*Anderson et al. 2005*).

## RESISTANCE TO RADIATION THERAPY

Radiation therapy is a traditional method of cancer treatment. Normal tissues are subjected to multiple doses of radiation therapy over a number of weeks to minimize damage to normal tissues and achieve therapeutic effects to the optimum. However, this method of giving interspersed radiation doses can be linked with accelerated repopulation, leading us to believe that a subset of radiation-resistant cells undergo proliferation. Experimental data from breast or brain cancer models indicate that CSC are endowed with resistance to radiation and suggest that CSC numbers in a tumor can affect relapse after radiotherapy (*Yin et al. 1997*). It was demonstrated in a glioblastoma model that the CD133+ fraction, enriched in CSC, is more resistant to radiation-induced apoptosis than its CD133- counter-part. The percentage of CD133+ cells following irradiation was increased both in vitro and in tumors (*Bao et al. 2006*). Experimental studies conducted with other groups showed in breast cancer cell lines a similar enrichment for the progenitor cell-containing SP after irradiation (*Chen et al. 2007*). An example of CSCs being provided with very efficient mechanisms of DNA damage repair, giving them the ability to activate DNA damage check points more readily than the rest of the cancerous cells is provided in glioblastoma cell lines. Here, the CD133+

subpopulation displays a basal activation of some proteins involved in the DNA damage check point, such as the checkpoint kinases Chk1/2. Inhibition of these kinases with a small molecule inhibitor disabled radioresistance of CSC-enriched cells (*Bao et al. 2006*). This efficient DNA damage repair is just one of many mechanisms that account for the particular resistance of CSC to radiation therapy.

## OVERCOMING CSC AND SP RESISTANCE TO CANCER THERAPEUTICS

According to the CSC hypothesis, CSC are more resistant to therapy than non-CSC. Thus, changes in tumor volume after therapy, which are governed by modification of the total burden of cancer cells, may not serve as an indicator of tumor eradication. It is absolutely essential to get rid of the sub-population of rare CSCs in order to effectively control certain tumours. As a result, fundamental optimization of cancer therapies is required in order to target CSC.

CSC and SP within a tumor can be targeted using two possible strategies:

- (i) introducing treatments able to target molecular pathways which are specifically over-activated in malignant stem cells
- (ii) sensitizing CSC to standard therapies by blocking those mechanisms which cause these cells to survive and as a result, lead to their resistance to cancer therapeutics.

## STEM CELLS FOR CANCER THERAPY

### Source:

The ideal choice of the source of stem cells for therapeutic purposes is ESCs. This is due to their higher totipotency and indefinite life span as compared to ASCs (adult stem cells), which have lower totipotency and restricted life span.

But, the use of ESCs have limitations which are mostly ethical in nature. As a result, their application in therapy and research is not without restrictions. Another cause for concern is that the stem cells with higher totipotency have been shown to be more tumorigenic in mice (*Serakinci et al. 2004*). This is why, due to their greater ease of availability and fewer ethical constraints, ASCs are the stem cells with wider

research and therapeutic applications. ASCs are also more easily accessible as compared to ESCs making them the preferred choice. The most commonly studied stem cells are ASCs from bone marrow (HSCs & MSCs). MSCs support HSCs in the bone marrow and have the ability to differentiate both in vivo and in vitro into the different mesenchymal cells such as bone, cartilage, fat, muscle, tendon and marrow stroma (Simonsen et al. 2002).

ESCs are obtained from 5-day old pre-implantation human embryos. ASCs can be obtained from many tissues including bone, synovium, deciduous teeth, adipose tissue, brain, blood vessels, blood and umbilical cord blood (Awad et al. 2004, Lee et al. 2004). While ethical and legal reasons restrict the use of ESCs in research and clinical fields, ASCs constitute the main source for the stem cells. These ASCs are mostly derived from the bone marrow and peripheral blood. One of the most common procedures performed to obtain ASCs is the bone marrow (BM) aspiration, but it is associated with morbidity in the form of wound infection and sepsis complications (Pittenger et al. 1999). ASCs can also be obtained from adipose tissues such as abdominal fat and infra-patellar fat (Huang et al. 2004). This is a less invasive and less morbid option than the bone marrow aspiration. Biological studies conducted have shown that there is no significant difference in the cell growth kinetics, cell senescence, gene transduction of adherent stromal cells and yield from stem cells obtained from bone marrow or adipose tissues (De Ugarte et al. 2003). The peripheral blood also provides a safe and easily accessible route for Another procedure involving minimal morbidity is by using ASCs through peripheral blood. The use of ASCs through peripheral blood has shown to induce more T and NK (Natural Killer) cells compared to bone marrow ASCs (Talmadge et al. 1997).

#### **CHOICE OF TYPE OF STEM CELL: BONE MARROW OR PERIPHERAL BLOOD**

The most commonly used stem cell sources are the bone marrow and the peripheral blood. The disadvantage of the bone marrow aspiration procedure lies in the fact that it is invasive and is associated with the potential possible complications which consist of fracture, wound infection and sepsis. On the other hand, the procedure for PBSCs isolation is not as invasive.

A greater number of CD4 T and NK cells are induced by PBSCs as compared to stem cells obtained from the bone marrow (Talmadge et al. 1997). As a result, the stem cells from peripheral blood are considered the preferred source of stem cells. However, it has been noted that the occurrence of graft versus host reaction varies with PBSCs compared to BM stem cells (Couban et al. 2003). Double stem cell transplantation has been documented to improve overall survival compared to single stem cell transplantation (Attal et al. 2003). It has been reported that Granulocyte-colony stimulating factor (G-CSF) helps in proliferation and differentiation of haematopoietic progenitor cells (Szyper-Kravitz et al. 2003). G-CSF has also been reported to mobilise autologous peripheral blood stem cells and to preserve and increase the length of telomerase (Szyper-Kravitz et al. 2003). A number of agents which are shown to enhance the G-CSF activity in mobilising stem cell are paclitaxel and docetaxel (Danova et al. 2000), recombinant human thrombopoietin (Somlo et al. 1999), lithium (Canales et al. 1999) and recombinant methionyl human stem cell factor (r-metHuSCF) (Prosper et al. 2003).

#### **STEM CELLS IN TISSUE REGENERATION AND AS DELIVERY VEHICLES**

Stem cells from haemopoietic tissues have dazzling properties that enable them to switch between haematopoietic and non-haematopoietic lineages, exhibiting an unexpected degree of developmental or differentiation potential. Needless to say, theoretically, this allows HSC to be used to regenerate any non-haematopoietic tissue (Martin-Rendon & Watt 2003). The applications of this technique lies in bone tumours as the reconstruction of bone following chemotherapy and surgery is always a problem that has to be dealt with efficiently. For instance, the stromal stem cells derived from bone marrow have been used in the cell-based bone reconstruction following chemotherapy and surgery in osteosarcoma and Ewing sarcoma (Becccheroni et al. 2003). The regeneration of osteoblasts from the survived mesenchymal progenitor cells following COSS-96 (the cooperative osteosarcoma study) polychemotherapy in vitro and its potential in vivo use has been shown by Jager et al. (2005). A number of clinical trials showing role of the stem cells in the regeneration of myocardial tissue following

myocardial infarction have been conducted (*Klein et al. 2007*). Systematic drug delivery or gene therapy are techniques showing promise, but is several factors such as immune detection, non-specific accumulation in normal tissues and poor permeation pose constraints to this procedure. It can also be said, that the effects of many anticancer agents are restricted due to either their toxicities or their short half lives such as interferon  $\beta$ , which shows anti-proliferative and pro-apoptotic activities in vitro, but has shown restricted effects on human malignancies in vivo (*Chawla-Sarkar et al. 2001*). A solution that can be proposed for these would be the cell-based carriers that may target the desired site.

The concept of use of stem cells as delivery vehicles owes its origin to the fact that tumors, acting like injuries, send out chemo-attractants such as the vascular endothelial growth factor (VEGF) to recruit MSC to form the supporting stroma of the tumor, and pericytes for angiogenesis. This MSC which is transduced with an adenoviral expression vector carrying interferon-P gene has been demonstrated to increase the production of interferon-P at the local site (*Studenly et al. 2004*). This in vivo function of MSC is dependent to an extent on signals from the target tissue microenvironment. An example that can be cited here is that the tissues such as skin would have high cell tum over where there would be more signals for MSC compared to connective tissues where the high cell turnover is apparent only during healing process (*Studenly et al. 2004*). In a similar fashion, MSC engineered to release interferon-P has been reported to create high local interferon-P levels in the mice glioma (*Nakamizo et al. 2005*). The neural stem cells have been reported as the delivery vehicles for the gene therapy for CNS disorders (*Muller et al. 2006*). *Liu et al. (2010)* demonstrated high-dose therapy/ autologous stem cell transplantation can improve cure rate and prolong survival time significantly in patients with nasopharyngeal T cell lymphomas (*Muller et al.*

*2006*). There also seems to be promise in the future for the use of the endothelial progenitor cells as the delivery vehicles for gene therapy because of their attraction towards the site of angiogenesis rather than the quiescent vasculature (*Anderson et al. 2005*). Stem cells can also be made to deliver immune-activating cytokines and other secreted proteins to brain and breast tumors (*Hayashi et al. 2011*).

## CONCLUSION

As our understanding of stem-cell behaviour rapidly increases, more and more reports suggest that use of stem-cell therapy will extend well beyond regenerative medicine in the near future. The traditional therapies of chemotherapy and radiotherapy often lead to initial shrinkage of tumours but, eventual development of metastatic drug-resistant disease in the treated patients. As a result, the use of stem cells in immuno-modulation or reconstitution is one of the most important methods in cancer therapy. Due to their inherent tumortropic migratory properties, stem cells can serve as vehicles for the delivery of effective, targeted treatment to isolated tumours and to metastatic disease. In vitro, stem cells can readily be engineered by inserting specifically tailored transgenes with antitumour effects to create tumour-seeking therapeutic vehicles. Transgene effects include direct tumour-cell killing, promotion of local immune responses, oncolytic virus production, and prodrug activation schemes. Stem cell transplants can be from the bone marrow, the peripheral blood and in case of children and young adults, from the umbilical cord blood which is stored, and frozen. The battle against cancer rages stronger than ever before. Advanced studies and new insights into avenues for stem-cell sourcing have shortened the probable time to realize the long awaited victory against cancer and to rescue millions of patients, their hopes and dreams from its curse.

## References

- Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF 2003, 'Prospective identification of tumorigenic breast cancer cells; Proc Natl Acad Sci USA, vol.100, pp.3983-3988.
- Anderson SA, Glod J, Arbab AS, Noel M, Ashari P, Fine HA, Frank JA 2005, 'Noninvasive MR imaging of magnetically labeled stem cells to directly identify neovasculature in a glioma model'. *Blood*, vol.105, pp.420-425.
- Attal M, Harousseau JL, Facon T, Guilhot F, Doyen C, Fuzibet JG, Monconduit M, Hulin C, Caillot D, Bouabdallah R, Voillat L, Sotto JJ, Grosbois B, Bataille R 2003, 'Single versus double autologous stem-cell transplantation for multiple myeloma'. *N Engl J Med*, vol.349, pp.2495-2502.
- Awad HA, Wickham MQ, Leddy HA, Gimble JM, Guilak F 2004, 'Chondrogenic differentiation of adipose-derived adult stem cells in agarose, alginate, and gelatin scaffolds; *Biomaterials*, vol. 25, pp.3211-3222.
- Bao S, Q Wu, R.E. Mclendon, Y. Hao, Q Shi, A.B. Hjelmeland, MW. Dewhirst, D.D. Bigner, J.N. Rich 2006, 'Glioma stem cells promote radioresistance by preferential activation of the DNA damage response; *Nature*, vol.444, pp.756-760.
- Beccheroni A, Lucarelli E, Donati D, Sangiorgi L, Capponcelli S, Gorini M, Zambon BA, Giardino R, Mercuri M, Ferrari S, Bacci G, Picci P 2003, 'Recovery of stromal stem cells in bone sarcoma patients after chemotherapy: implication for cell-based therapy in bone defect reconstruction'. *Oncol Rep*, vol. 10, pp.891-896.
- Bidlingmaier S, Zhu X, Liu B 2008, 'The utility and limitations of glycosylated human CD133 epitopes in defining cancer stem cells'. *J Mol Med*, vol.86, pp.1025
- Canales MA, Arrieta R, Hernandez-Garcia C, Bustos JG, Aguado MJ, Hernandez-Navarro F 1999, 'A single apheresis to achieve a high number of peripheral blood CD34+ cells in a lithium-treated patient with acute myeloid leukaemia' *Bone Marrow Transplant*, vol.23, pp.305.
- Chawla-Sarkar M, Leaman OW, Borden EC 2001, 'Preferential induction of apoptosis by interferon (IFN)-beta compared with IFN-alpha2: correlation with TRAIUApo2L induction in melanoma cell lines; *Clin Cancer Res*, vol. 7, pp.1821-1831.
- Chen MS, WA. Woodward, F. Behbod, S. Peddibhotla, MP. Alfaro, T.A. Buchholz, J.M. Rosen 2007, 'Wnt/beta-catenin mediates radiation resistance of Sea 1+ progenitors in an immortalized mammary gland cell line; *J. Cell Sci.* vol.120, pp. 468-477.
- Chiba Tetsuhiro, Kamiya Akihide, Yokosuka Osamu, Iwama Atsushi 2009, 'Cancer stem cells in hepatocellular carcinoma: Recent progress and perspective; *Cancer Letters*, vol.286, pp.145-153.
- Chua C, N Zaiden, K.H. Chong, S.J. See, M.C. Wong, B.T. Ang, C. Tang 2008, 'Characterization of a side population of astrocytoma cells in response to temozolomide; *J. Neurosurg*, vol.109 pp. 856-866.
- Clarke Michael F 2005, 'Self-Renewal and Solid-Tumor Stem Cells; *Biology of Blood and Marrow Transplantation*, vol.11, pp.14-16.
- Collins AT, Berry PA, Hyde C, Stower MJ, Maitland NJ. 2005, 'Prospective identification of tumorigenic prostate cancer stem cells'. *Cancer Res*, vol.65, pp.10946-10951.
- Couban S, Simpson DR, Barnett MJ, Bredeson C, Hubesch L, Howson-Jan K, Shore TB, Walker IR, Browett P, Messner HA, Panzarella T, Lipton JH 2002, 'A randomized multicenter comparison of bone marrow and peripheral blood in recipients of matched sibling allogeneic transplants for myeloid malignancies'. *Blood*, vol. 100, pp.1525-1531
- Daniel WW, Brunschwig A. 1961, 'The management of recurrent carcinoma of the cervix following simple total hysterectomy'. *Cancer*, vol.14, pp.582-586.
- Danova M, Porta C, Ferrari S, Brugnattelli S, Comolli G, Riccardi A, Ascari E 2000, 'Cell cycle status and apoptosis of hematopoietic progenitor cells released into the peripheral blood after taxanes and granulocyte colony-stimulating factor in breast cancer patients'. *Oncol Rep*, vol.7, pp.585-589.



- De Ugarte DA, Morizono K, Elbarbary A, Alfonso Z, Zuk PA, Zhu M, Dragoo JL, Ashjian P, Thomas B, Benhaim P, Chen J, Fraser J, Hedrick MH 2003, 'Comparison of multi-lineage cells from human adipose tissue and bone marrow'. *Cells Tissues Organs*, vol.174, pp.101-109.
- Dick JE. 1996, 'Normal and leukemic human stem cells assayed in SCID mice'. *Semin Immunol*, vol.8, pp.197-206.
- Eramo A, Lotti F, Sette G, Pilozi E, Biffoni M, Di Virgilio A. 2008, 'Identification and expansion of the tumorigenic lung cancer stem cell population'. *Cell Death Differ*, vol.15, pp.504- 514.
- Fang D, Nguyen TK, Leishear K, Finko R, Kulp AN, Hotz S, et al. 2005, 'A tumorigenic subpopulation with stem cell properties in melanomas'. *Cancer Res*, vol.65, pp.9328-9337.
- Gibbs CP, Kukekov V, Reith JD, Tchigrinova O, Suslov ON, Scott EW, et al. 2005, 'Stem-like cells in bone sarcomas: implications for tumorigenesis'. *Neoplasia*, vol.7, pp.967-976.
- Harrington L. 2004, 'Does the reservoir for self-renewal stem from the ends?' *Oncogene*, vol.3, pp.7283-7289.
- Hayashi S, Fujita K, Matsumoto S, Akita M, Satomi A 2011, 'Isolation and identification of cancer stem cells from a side population of a human hepatoblastoma cell line, HuH-6 clone-5'. *Pediatr Surg Int*. vol. 27, pp.9-16.
- Hirschmann-Jax C, A.E. Foster, G.G. Wulf, J.G. Nuchtern, T.W. Jax, U.Gobel, M.A. Goodell, M.K. Brenner 2004, 'A distinct "side population" of cells with high drug efflux capacity in human tumor cells; *Proc. Natl. Acad. Sci. USA*, vol. 101, pp.14228-14233.
- Houghton JeanMarie , Alexei Morozov b, Iva Smirnova , Timothy C. Wang 2009, 'Stem cells and cancer; *Seminars in Cancer Biology* ,vol.17, pp.191-203.
- Huang JI, Zuk PA, Jones NF, Zhu M, Lorenz HP, Hedrick MH, Benhaim P 2004, 'Chondrogenic potential of multipotential cells from human adipose tissue'. *Plast Reconstr Surg*, vol.113, pp.585-594.
- Jager M, Schultheis A, Westhoff B, Krauspe R 2005, 'Osteogenic progenitor cell potency after high-dose chemotherapy (COSS-96)'. *Anticancer Res*, vol.25, pp.947-954.
- Klein HM, Ghodsizad A, Marktanner R, Poll L, Voelkel T, Mohammad Hasani MR, Piechaczek C, Feifel N, Stockschlaeder M, Burchardt ER, Kar BJ, Gregoric J, Gams E 2007, 'Intramyocardial implantation of CD133+ stem cells improved cardiac function without bypass surgery'. *Heart Surg Forum*, vol.10, pp.E66-E69.
- Krishnamurthy P, D.D. Ross, T. Nakanishi, K. Bailey-Dell, S. Zhou, K.E. Mercer, B. Sarkadi, B.P. Sorrentino, J.D. Schuetz 2004, 'The stem cell marker Bcrp/ABCG2 enhances hypoxic cell survival through interactions with heme; *J. Biol. Chem.*vol.279, pp.24218-24225.
- Lanza F, Healy L, Sutherland DR 2001, 'Structural and functional features of the CD34 antigen: an update; *J Biol Regul Homeost Agents*, vol.15, pp.1-13.
- Lee OK, Kuo TK, Chen WM, Lee KD, Hsieh SL, Chen TH 2004, 'Isolation of multipotent mesenchymal stem cells from umbilical cord blood; *Blood*, vol.103, pp.1669-1675.
- Li L, Borodyansky L, Yang Y 2009. 'Genomic instability en route to and from cancer stem cells; *Cell Cycle* 8:1000-1002
- Liu XM, Zhang HL, Cui XZ, Wang HQ 2010, 'High-dose chemotherapy/autologous stem cell transplantation for nasopharyngeal T cell lymphoma; *Zhonghua Xue Ye Xue Za Zhi*, vol.31, no.8, pp.515-518.
- Lobo NA, Shimono Y, Qian D, Clarke MF 2007, 'The biology of cancer stem cells; *Annu Rev Cell Dev Biol*, vol.23, pp.675-699.
- Martin C.M., A. Ferdous, T. Gallardo, C. Humphries, H. Sadek, A. Caprioli, J.A. Garcia, L.I. Szveda, M.G. Garry, D.J. Garry 2008, 'Hypoxia-inducible factor-2 $\alpha$  transactivates *Abcg2* and promotes cytoprotection in cardiac side population cells; *Circ. Res.*, vol.102, pp.1075-1081.

- Martin C.M., A. Ferdous, T. Gallardo, C. Humphries, H. Sadek, A. Caprioli, J.A. Garcia, L.I. Szweda, M.G. Garry, D.J. Garry 2008, 'Hypoxia-inducible factor-2 $\alpha$  transactivates Abcg2 and promotes cytoprotection in cardiac side population cells; *Circ. Res.*, vol.102, pp.1075-1081.
- Martin-Rendon E, Watt SM 2003, 'Exploitation of stem cell plasticity; *Transfus Med*, vol.13, pp. 325-349.
- Millera Stanley J, Robert M. Lavker, Tung-Tien Sun 2005, Interpreting epithelial cancer biology in the context of stem cells: Tumor properties and therapeutic implications, *Biochimica et Biophysica Acta*, pp. 1756 25 - 175652.
- Mizrak D, Brittan M, Alison MR 2008, 'CD133: molecule of the moment; *J Pathol*, vol.214, pp.3-9.
- Moserle Lidia, Margherita Ghisi, Alberto Amadori, Stefano Indraccolo 2010, 'Side population and cancer stem cells: Therapeutic implications; *Cancer Letters*, vol. 288, pp. 1-89.
- Muller FJ, Snyder EY, Loring JF 2006, 'Gene therapy: can neural stem cells deliver?' *Nat Rev Neurosci*, vol.?, pp.75-84.
- Nakamizo A, Marini F, Amano T, Khan A, Studeny M, Gumin J, Chen J, Hentschel S, Vecil G, Dembinski J, Andreeff M, Lang FF 2005, 'Human bone marrow-derived mesenchymal stem cells in the treatment of gliomas; *Cancer Res*, vol. 65, pp.3307-3318.
- Piechaczek C 2001, '(0133; *J Biol Regul Homeost Agents*, vol.15, pp.101-102.
- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti OW, Craig S, Marshak DR 1999, 'Multilineage potential of adult human mesenchymal stem cells'. *Science*, vol.284, pp.143-147.
- Potgens AJ, Bolte M, Huppertz B, Kaufmann P, Frank HG 2001, 'Human trophoblast contains an intracellular protein reactive with an antibody against CD133 - a novel marker for trophoblast; *Placenta*, vol.22, pp.639-645.
- Prosper F, Sola C, Hornedo J, Arbona C, Menendez P, Orfao A, Lluch A, Cortes-Funes H, Lopez JJ, Garcia-Conde J 2003, 'Mobilization of peripheral blood progenitor cells with a combination of cyclophosphamide, r-metHuSCF and filgrastim in patients with breast cancer previously treated with chemotherapy; *Leukemia*, vol.17, pp.437-441.
- Reya T, Morrison SJ, Clarke MF, Weissman IL 2001, 'Stem cells, cancer, and cancer stem cells; *Nature*, vol.414, pp.105-111.
- Richardson GD, Robson CN, Lang SH, Neal DE, Maitland NJ, Collins AT 2004, 'CD133, a novel marker for human prostatic epithelial stem cells; *J Cell Sci*, vol.117, pp.3539-3545.
- Ryan DH, Chapple CW, Kossover SA, Sandberg AA, Cohen HJ 1987, 'Phenotypic similarities and differences between CALLA-positive acute lymphoblastic leukemia cells and normal marrow CALLA-positive B cell precursors; *Blood*, vol.70, pp. 814-821.
- Salven P, Mustjoki S, Alitalo R, Alitalo K, Rafii S 2003, 'VEGFR-3 and CD133 identify a population of CD34+ lymphatic/vascular endothelial precursor cells; *Blood*, vol.101, pp.168-172.
- Sanaei N, Alvarez-Buylla A, Berger MS 2005, 'Neural stem cells and the origin of gliomas; *N Engl J Med*, vol. 353, pp.811-822.
- Schofield R 1978, 'The relationship between the spleen colony-forming cell and the haemopoietic stem cell; *Blood Cells*, vol.4, pp.7-25.
- Serakinci N, Guldberg P, Burns JS, Abdallah B, Schroder H, Jensen T, Kassem M 2004, 'Adult human mesenchymal stem cell as a target for neoplastic transformation; *Oncogene*, vol.23, pp.5095-5098.
- Simonsen JL, Rosada C, Serakinci N, Justesen J, Stenderup K, Rattan SI, Jensen TG, Kassem M 2002, 'Telomerase expression extends the proliferative life-span and maintains the osteogenic potential of human bone marrow stromal cells; *Nat Biotechnol*, vol.20, pp.592-596.

- Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, et al. 2003, 'Identification of a cancer stem cell in human brain tumors' *Cancer Res*, vol.63, pp.5821-5828.
- Singh SK, Hawkins C, Clarke ID, Squire JA, Bayani J, Hide T, et al. 2004, 'Identification of human brain tumour initiating cells; *Nature*, vol.432, pp.396-401.
- Sneddon JB, Werb Z 2007, 'Location, location, location: the cancer stem cell niche; *Cell Stem Cell*, vol.1, pp.607-611.
- Som lo G, Sniecinski J, ter Veer A, Long mate J, Knutson G, Vuk-Pavlovic S, Bhatia R, Chow W, Leong L, Morgan R, Margolin K, Raschko J, Shibata S, Tetef M, Yen Y, Forman S, Jones D, Ashby M, Fyfe G, Hellmann S, Doroshow JH 1999, 'Recombinant human thrombopoietin in combination with granulocyte colony-stimulating factor enhances mobilization of peripheral blood progenitor cells, increases peripheral blood platelet concentration, and accelerates hematopoietic recovery following high-dose chemotherapy; *Blood*, vol.3, pp.2798-2806
- Stern R 2008, 'Association between cancer and "acid mucopolysaccharides": an old concept comes of age, finally; *Semin Cancer Biol*, vol.18, pp.238-243.
- Studeny M, Marini FC, Champlin RE, Zompetta C, Fidler IJ, Andreeff M 2002, 'Bone marrow-derived mesenchymal stem cells as vehicles for interferon-beta delivery into tumors; *Cancer Res*, vol.2, pp.3603-3608.
- Studeny M, Marini FC, Dembinski JL, Zompetta C, cabreira-Hansen M, Bekele BN, Champlin RE, Andreeff M 2004, 'Mesenchymal stem cells: potential precursors for tumor stroma and targeted-delivery vehicles for anticancer agents; *J Natl Cancer Inst*, vol.96, pp.1593-1603.
- Szotek PP, R Pieretti-Vanmarcke, PT, Masiakos, D.M. Dinulescu, D.Connolly, R Foster, D. Dombkowski, F. Preffer, D.T. Maclaughlin, P.K. Donahoe 2006, 'Ova\_rian cancer side population defines cells with stem cell-like characteristics and mullerian inhibiting substance responsiveness; *Proc. Natl. Acad. Sci. USA*, vol.103, pp.11154-11159.
- Szyper-Kravitz M, Uziel O, Shapiro H, Radnay J, Katz T, Rowe JM, Lishner M, Lahav M 2003, 'Granulocyte colony-stimulating factor administration upregulates telomerase activity in CD34+ haematopoietic cells and may prevent telomere attrition after chemotherapy; *Br J Haematol*, vol.120, pp.329-336.
- Talmadge JE, Reed E, Ino K, Kessinger A, Kuszynski C, Heimann D, Varney M, Jackson J, Vose JM, Bierman PJ 1997, 'Rapid immunologic reconstitution following transplantation with mobilized peripheral blood stem cells as compared to bone marrow; *Bone Marrow Transplant*, vol.19, pp.161-172.
- Martin-Rendon E, Watt SM 2003, 'Exploitation of stem cell plasticity; *Transfus Med*, vol. 13, pp.325-349.
- Uchida N, Buck DW, He D, Reitsma MJ, Masek M, Phan TV, et al. 2000, 'Direct isolation of human central nervous system stem cells; *Proc Natl Acad Sci USA*, vol.97, pp.14720-14725.
- Vormoor J, Lapidot T, Ptlumio F, Risdon G, Patterson B, Broxmeyer HE, et al. 1994, 'Immature human cord blood progenitors engraft and proliferate to high levels in severe combined immunodeficient mice'. *Blood*, vol.83, pp.2489-2497.
- Wang Jean C.Y. and John E Dick 2005, 'Cancer stem cells: Lessons from leukemia; *Trends in Cell Biology*, vol.15, no.9, pp.494-501.
- Yang YM, Chang JW 2008, 'Current status and issues in cancer stem cell study; *Cancer Invest*, vol. 26, pp.741-755.
- Yin AH, Miraglia S, Zanjani ED, Almeida-Porada G, Ogawa M, Leary AG, et al. 1997, 'AC133, a novel marker for human hematopoietic stem and progenitor cells; *Blood*, vol.90, pp.5002-5012.
- Yu Y, Flint A, Dvorin EL, Bischoff J 2002, 'AC133-2, a novel isoform of human AC133 stem cell antigen; *J Biol Chem*.vol.277, pp.20711-20716.